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Abstract

Magnetic iron oxide nanoparticles (IONs) stand out among a plethora of drug nanocarriers as sturdy nanoplatfoms due to exceptional magnetic and biological properties, which allow them to achieve significant drug loading as well as targeting capabilities. These applications necessitate accurate nanoparticle design in terms of numerous characteristics that must be evaluated in tandem to achieve maximum therapeutic efficacy. This concise overview summarizes recent advances in the roles of untreated and modified iron oxide nanoparticles for drug delivery. These modifications included chitosan, poly(vinylpyrrolidone), poly(vinyl alcohol), poly(lactic-co-glycolic acid), and poly(ethylene glycol). One of the key areas of research in the targeted drug delivery domain is the invention of nanocarriers that allow for the efficient delivery of therapeutic chemicals to specific sites. Drugs loaded onto iron oxide nanoparticles can be efficiently guided and selectively delivered to selected sites by precisely altering the structural features of the nanoparticles.

Keywords
(separated by '-')

Iron oxide - Nanostructures - Drug delivery - Magnetic

Application of Magnetic Iron Oxide Nanostructures in Drug Delivery: A Compact Review



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Abstract Magnetic iron oxide nanoparticles (IONs) stand out among a plethora of drug nanocarriers as sturdy nanoplatforms due to exceptional magnetic and biological properties, which allow them to achieve significant drug loading as well as targeting capabilities. These applications necessitate accurate nanoparticle design in terms of numerous characteristics that must be evaluated in tandem to achieve maximum therapeutic efficacy. This concise overview summarizes recent advances in the roles of untreated and modified iron oxide nanoparticles for drug delivery. These modifications included chitosan, poly(vinylpyrrolidone), poly(vinyl alcohol), poly(lactic-co-glycolic acid), and poly(ethylene glycol). One of the key areas of research in the targeted drug delivery domain is the invention of nanocarriers that allow for the efficient delivery of therapeutic chemicals to specific sites. Drugs loaded onto iron oxide nanoparticles can be efficiently guided and selectively delivered to selected sites by precisely altering the structural features of the nanoparticles.

Keywords Iron oxide · Nanostructures · Drug delivery · Magnetic

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1

15 Introduction

16 Nanostructures have developed over the past few years into new, potent tools in
17 a variety of technical applications [1–10]. Researchers have become increasingly
18 interested in these applications, which has led to excellent developments in the
19 creation of many types of nanomaterials and nanodevices [11–17]. Due to their
20 inherent magnetic properties (superparamagnetism), magnetic iron oxide nanoparti-
21 cles (IOMNPs) have received the most attention among the various types of nano-
22 materials studied [18, 19]. These properties allow them to be used in a variety of
23 scientific fields like electronics or the environment. Ions are the perfect platform
24 for biological applications because of their exceptional magnetic characteristics as
25 well as their biocompatibility, stability, and environmental friendliness [20]. Co-
26 precipitation, sol–gel, hydrothermal, and other techniques are used to create magnetic
27 nanoparticles, among others [18]. The co-precipitation approach may be used to
28 create magnetite nanoparticles with a narrow size distribution and a particle size
29 range from 5 to 100 nm, and the magnetism of the nanoparticles can be improved by
30 clustering them [18, 19].

31 Targeted nanomedicine using magnetic nanoparticles has emerged as one of the
32 most potent methods for diagnosis and therapy [21]. One more benefit is the ability to
33 employ an external magnet to deliver medications to target organs utilizing the core
34 magnetic nanoparticle. This could solve a number of issues, including off-target
35 adverse effects, poor drug solubility, brief life cycle, etc. [22]. Due to changes in
36 the drug’s pharmacokinetics, an extension of the drug cycle’s half-life, a consistent
37 release of the medication, and a decrease in the toxicity associated with the drug, drug
38 delivery systems based on nanotechnology have significantly improved therapeutic
39 outcomes [23, 24]. Cell separation, immunoassay, tissue regeneration, hyperthermia,
40 tumor targeting, and drug administration are just a few of the uses for inorganic
41 magnetic iron oxide nanoparticles (MNPs), which have the generic chemical formula
42 Fe_3O_4 . Due to their particular characteristics, which include small particle size,
43 superparamagnetic and specific magnetic features, low toxicity, high half-life, and
44 catalytic activities, MNPs have come to be the focus of innovative materials science
45 [25–27].

46 Magnetic iron oxide nanostructures with a variety of capabilities could acquire
47 desirable characteristics by adding functional groups to their surfaces [25]. For
48 instance, magnetic iron oxide nanoparticles can be coated with many active
49 medications as well as other active ingredients, opening up a wide range of
50 possible uses in nanomedicine, such as targeted drug delivery. These nanoparti-
51 cles, which can migrate toward magnets, are being researched as potential medica-
52 tion delivery vehicles. In this work, the roles of untreated and modified iron oxide
53 nanoparticles for drug delivery were investigated. These modifications included
54 chitosan, poly(vinylpyrrolidone), poly(vinyl alcohol), poly(lactic-co-glycolic acid),
55 and poly(ethylene glycol).

Drug Delivery

As new materials and technologies were developed, there was a surge in the study and development of therapeutic drug delivery systems. More sophisticated pharmaceutical systems were made possible by the advancement of biotechnology and our understanding of physiological mechanisms. As a result, there has been an increase in drug delivery system development strategies that exhibit improved abilities to alter and regulate the administration of active substances [28, 29]. Nanotechnology is one of the most often used methods in this area for regulating drug distribution and improving the effectiveness, safety, and caliber of the systems. In addition, patient therapy is enhanced. Iron oxide nanoparticles (IOMNPs) are useful for regulating drug distribution and for enabling drug targeting, one of the most crucial tactics [29]. These nanoparticles and/or implantable magnets can be used in magnetic drug targeting, which involves delivering the particles to the desired place, fixing them there while the active substance is released, and acting locally [29]. This method can lower dosage requirements and get rid of unwanted effects. IOMNPs are undergoing trials to look into the possibility that they could be used as drug carriers in this situation. The focus of Turrina et al. (2021) is on the binding patterns of the bee venom peptide lasioglossin III on bare IOMNPs [32]. Due to its cationic characteristics and strong binding potential, lasioglossin has a strong antibacterial behavior. The maximum drug loading of 22.7% is reached in phosphate-buffered saline when the effects of pH, buffer type, particle concentration, and time are taken into account. Temperature and salt content were shown to be sensitive desorption conditions after analysis. Using dynamic light scattering, zeta potential, and infrared spectroscopy, the nanoparticles and peptide-ion complexes are examined. Furthermore, cytotoxicity tests done on *Escherichia coli* reveal that bound lasioglossin has greater antibacterial action than free peptide. For the development of cationic peptide drug delivery carriers, bare IOMNPs are a promising platform material. The biocompatibility of Fe₃O₄ NPs was examined by Kansara et al. [33] using cytotoxicity assays and cell cycle analysis in the human breast adenocarcinoma cell line (MCF-7) as part of another study. Fe₃O₄ NPs were produced using the co-precipitation method. After 24 h of exposure to the two higher concentrations, flow cytometric analysis showed a significant ($P = 0.05$) increase in the internalization of Fe₃O₄ NPs in MCF-7 cells, as shown by an increase in the side scatter intensity. Fe₃O₄ NPs were shown to be biocompatible in the cytotoxicity experiments, NRU and MTT, since there was no significantly higher NRU (88% at concentration 150 M/mL) and a reduction in mitochondrial succinate dehydrogenase activity (96%) was detected at the maximum concentration after 24 h of exposure (Fig 2a, b). After a 24 h exposure, Fe₃O₄ NPs-treated MCF-7 cells showed no change in cell cycle progression. The results of the study showed that, despite being significantly internalized, Fe₃O₄ NPs synthesized using the safe-by-design method had no negative effects on cells as determined by cytotoxicity assays and cell cycle analysis in MCF-7 cells. As a result, these NPs may be used as a vehicle for the delivery of specific medications.

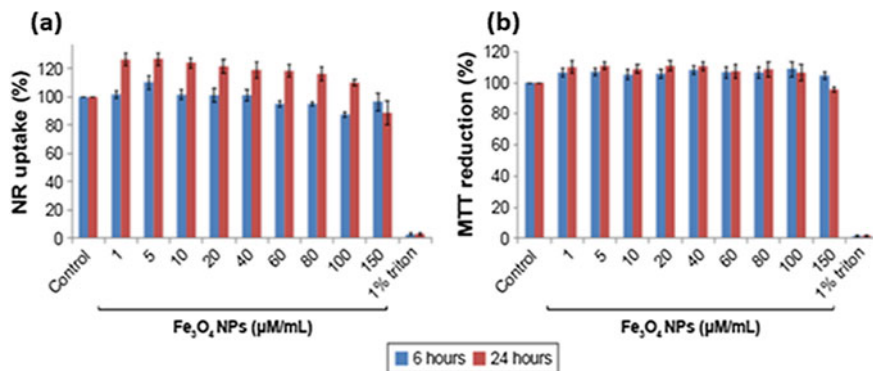


Fig. 1 Cytotoxicity of Fe₃O₄ NPs in MCF-7 cells. **a** NR uptake (%); **b** MTT reduction (%). Notes: The viability of the control cells was considered as 100%. Data are expressed as mean \pm standard error of the mean from three independent experiments. Abbreviations: NPs, nanoparticles; MCF-7, human breast adenocarcinoma cell line; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; NR, neutral red [33]

98 Polymeric layers, capsules, particles, or vesicles were suggested since the compo-
 99 sition of the exterior coating has a significant impact on the characteristics of these
 100 nanoparticles and their ability to successfully carry active compounds [28]. In order
 101 to make these particles biocompatible and appropriate for further functionalization
 102 through the attachment of various bioactive molecules, the surfaces of these parti-
 103 cles are often modified utilizing organic polymers and inorganic metals or oxides
 104 [34]. However, in order for the drug delivery system incorporating IOMNPs to be
 105 successful, a number of crucial factors must be taken into account.

106 The delivery system must have functional groups that can be further changed to
 107 control drug release or bind targeted units, as well as the ability to easily disperse in
 108 aqueous solutions [28] (Fig. 1).

109 To attach various medications to IOMNPs, nanostructured devices with a core-
 110 shell architecture are frequently used. The shell is the surface coating for nanoparticle
 111 functionalization, and the core is made up of nanoparticles. With this approach, the
 112 system's biodistribution, biocompatibility, and pharmacokinetics can all be enhanced
 113 [35]. As a result of their ability to shield nanoparticles from oxidation and give them
 114 stability, synthetic and natural polymers are the most often employed surface coatings
 115 in IOMNPs.

116 The following substances are used: poly(vinylpyrrolidone), polyvinyl alcohol,
 117 poly(lactic-co-glycolic acid), and chitosan [28]. Due to its non-fouling qualities and
 118 decreased opsonization of blood proteins, PEG—which is hydrophilic, uncharged,
 119 and biocompatible—was used to coat the IOMNPs. Due to their ability to avoid

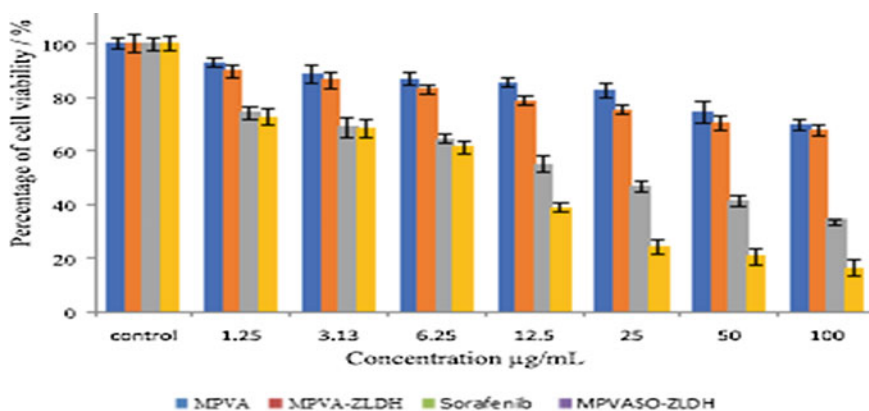


Fig. 2 Cytotoxicity assay of MPVA, MPVA-ZLDH (the nanocarriers), sorafenib, and MPVASO-ZLDH (the nanoparticles) against HepG2 cells at 72 h of incubation [25]

120 immune system detection, nanoparticles can prolong their duration in the blood-
 121 stream and accumulate in target cells and tissues [28]. Yallapu et al. (2010) investi-
 122 gated the potential of polyethylene glycol (PEG) functionalized magnetic nanopar-
 123 ticles (MNPs) as a drug delivery system, as an MRI agent, and as an antibody conju-
 124 gate [36]. A water-dispersible MNP formulation is created by coating an iron oxide
 125 core first with oleic acid (OA), then with OA-PEG. For prolonged drug delivery,
 126 hydrophobic doxorubicin partitions into the OA layer. The circulation of the MNPs
 127 was assessed in mouse carotid arteries, and the T1 and T2 MRI contrast qualities
 128 were established in vitro. The amine functional group on antibodies was coupled
 129 to an N-hydroxysuccinimide group (NHS) on the OA-PEG-80 for active targeting
 130 of the human MCF-7 breast cancer cell line. Results The optimized formulation
 131 comprised an iron-oxide core that was around 8 nm thick and a mean hydrody-
 132 namic diameter of 184 nm. The MNPs prolong the circulation period in vivo and
 133 reach a 30% relative concentration 50 min after injection. They also improve the T2
 134 MRI contrast. In vitro, doxorubicin-loaded MNPs exhibited sustained drug release
 135 and dose-dependent antiproliferative effects; the drug effect was boosted by trans-
 136 ferrin antibody-conjugated MNPs. The authors came to the conclusion that targeted
 137 medication delivery systems and MRI contrast agents could be created using PEG-
 138 functionalized MNPs. Mohanta et al. (2018) created almost spherical Fe₃O₄ nanopar-
 139 ticles with sizes in the 8–20 nm range using a sonication assisted co-precipitation
 140 approach to address the growing interest in magnetic nanoparticles for drug delivery
 141 applications [37]. The produced nanoparticles were further stabilized by coating
 142 with PEG-400, a hydrophilic and biocompatible polymer, to improve surface func-
 143 tioning. To assess their feasibility for drug delivery applications, various approaches
 144 were used to characterize both uncoated and PEG-400 coated iron oxide particles.
 145 The water-soluble anticancer medication daunorubicin hydrochloride was loaded
 146 onto the PEG-400 coated iron oxide nanoparticle for this purpose. Specifically, they
 147 changed the PEG/iron oxide ratio to optimize the PEG coating, which improved drug

148 loading and stability by effectively functionalizing the surface. In physiological and
149 acidic pH circumstances, the drug release study has been seen. A higher amount of
150 drug release at low pH indicated the functionalized particles' potential utility as drug
151 delivery vehicles. The functionalized particles also displayed pH-responsive drug
152 release.

153 Additionally, water-soluble synthetic polymers, polyvinylpyrrolidone (PVP) and
154 poly(vinyl alcohol) (PVA), are frequently employed in medicinal applications. The
155 preparation of hydrogel structures is made possible by their emulsifying and adhe-
156 sive characteristics. IOMNPs may participate in hydrogen bonding between polymer
157 chains, which prevents nanoparticle aggregation [30]. For instance, magnetic drug
158 nanoparticles made using (PVA) polymer, layered double hydroxide (LDHs), and
159 drug as the coating agent and magnetic iron oxide nanoparticles (MNPs) as the core
160 [25] were the subject of an investigation by Ebadi et al. in 2020. First, the Fe_3O_4
161 nanoparticles were created using the co-precipitation process. After that, Zn/Al-LDH,
162 sorafenib, and polyvinyl alcohol were applied to the surface (SO). Studies using XRD
163 and FTIR show that the iron oxide crystal structure is present in the core. The TGA
164 findings confirmed the existence of the core and shell. When the Fe_3O_4 nanopar-
165 ticles were coated with polyvinyl alcohol, Zn/Al-LDH, and the medication sorafenib,
166 the saturation magnetization was observed to be decreased from 80 to 57 emu/g.
167 According to HRTEM photographs, the average size of naked Fe_3O_4 nanoparticles
168 is about 30 nm. Additional structural analyses revealed that the addition of the shell
169 produced homogeneous particles with a particle size distribution of roughly 95 nm. It
170 was discovered that the pseudo-second-order equation controlled the kinetics of drug
171 release from the nanoparticles. The results of cell viability assays clearly demon-
172 strated that the magnetic iron oxide nanoparticles coated with polyvinyl alcohol-
173 sorafenib-Zn/Al-layered double hydroxide were more effective than sorafenib alone
174 against HepG2 liver cancer cells (Fig. 2), while they showed no cytotoxicity toward
175 3T3 fibroblasts (Fig. 3). These findings demonstrate the potential of the coated Fe_3O_4
176 magnetite nanoparticles as a drug delivery vehicle for biomedical applications.

177 The utilization of a polyglycolic acid and polylactic acid copolymer (PGA) in drug
178 delivery systems showed considerable promise [38]. This polymer may be formed
179 into many forms and sizes and is soluble in the majority of common solvents, making
180 it possible to encapsulate a variety of molecules [30]. As one method of maintaining
181 the particles in the joint cavity by an external magnetic field, and managing the
182 drug release for the treatment of arthritis and osteoarthritis, PLGA microparticles
183 comprising co-encapsulated dexamethasone acetate and IOMNPs were devised [39].
184 The ability of the DU145 prostate carcinoma cell line to proliferate in monolayer
185 culture was examined by Hajikarimi et al. (2014) for the uptake and cytotoxic effects
186 of magnetic poly lactic-co-glycolic acid (PLGA)-coated iron oxide nanoparticles as
187 a carrier of 5-fluorouracil (5-FU) and X-ray [40]. Following monolayer cultivation,
188 DU 145 cells were exposed to 2 Gy X-rays (6 Megavolts (MV)) and various doses of
189 5-FU or 5-FU-loaded nanoparticles for 24 h. Atomic adsorption spectroscopy was
190 then used to determine the rate of nanoparticle penetration (AAS). Using a colony

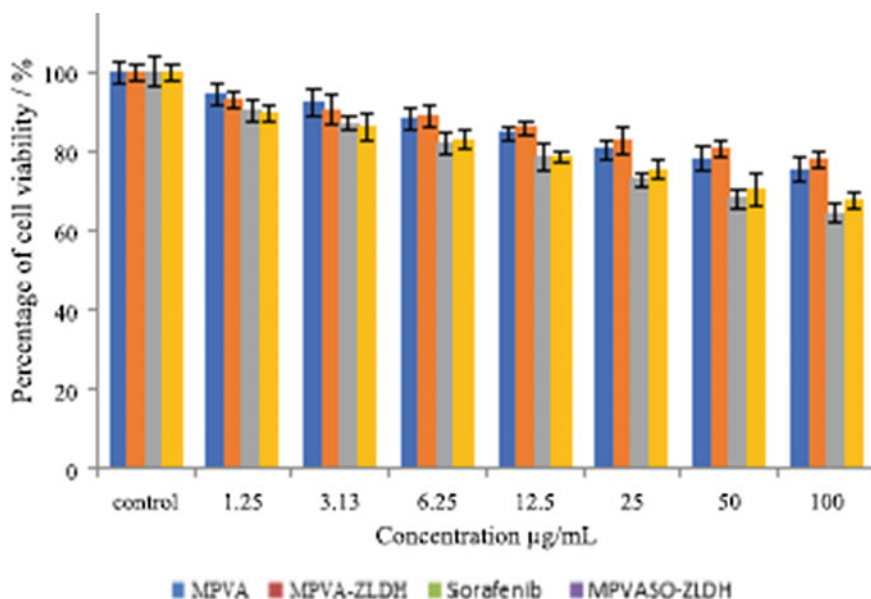


Fig. 3 Cytotoxicity assay of MPVA, MPVA-ZLDH (the nanocarriers), sorafenib, and MPVASO-ZLDH (the nanoparticles) against normal 3T3 cells at 72 h [25]

191 formation experiment, the cytotoxicity effect of these nanoparticles with/without X-
 192 ray radiation was assessed. Results from spectroscopy revealed that when nanopar-
 193 ticle concentrations increased, the iron content and thus the cellular absorption of
 194 5-FU-loaded nanoparticles increased. Additionally, as the quantities of 5-FU and 5-
 195 FU-loaded nanoparticles increased in combination with X-ray radiation, the ability
 196 of the cells to proliferate diminished. However, compared to free 5-FU, the amount
 197 of colony number reduction after treatment with 5-FU-loaded nanoparticles and 2 Gy
 198 of megavoltage X-ray radiation was much greater. Drug-loaded nanoparticles could
 199 therefore transport 5-FU into cells more effectively. Therefore, PLGA-coated iron
 200 oxide nanoparticles are efficient 5-FU drug delivery systems. The PLGA coating on
 201 iron oxide nanoparticles makes them biocompatible and provides a suitable surface
 202 for cell penetration.

203 The biodegradable polymer poly (lactic-co-glycolic acid, or PLGA), which
 204 exhibits appealing properties and provides opportunities to tailor the physicochem-
 205 ical properties, drug release properties, and biological behavior of the PLGA-
 206 based nanospheres, is widely used to create polymeric biodegradable carriers [29].
 207 When it comes to encapsulating highly hydrophilic medications like doxorubicin,
 208 double emulsion solvent evaporation techniques fall short (Dox). Before the PLGA
 209 nanospheres solidified, the drug diffused quickly into the exterior aqueous phase,
 210 which is the cause of this problem. Superparamagnetic iron oxide nanoparticles
 211 (SPIONs) and Dox were co-encapsulated with the PLGA polymer in a study by

Mosafer et al. (2018), with a focus on the double emulsion solvent evaporation-based methodologies to achieve the effective method in terms of nanoparticle size, particle size distribution, and drug loading [41]. The low affinity of Dox for the PLGA polymer could lead to limited entrapment efficiency because Dox has a high solubility in water. The chloroform solution was used to extract Dox into this organic phase with TEA to neutralize Dox in favor of Dox lipophilicity, improving the entrapment efficiency and LC. To improve the antiproliferation action of Dox inside cellular organelles like lysosomes (pH is approximately 5.5), faster Dox release of NPs at low pHs might be advantageous while restricting its release in blood circulation. In vitro Dox release from NPs in pH 7.4 PBS and pH 5.5 acetate buffers at 37 °C is depicted in Fig. 4. At pH 5.5, compared to pH 7.4, the cumulative Dox emission increased by almost 2.5 times. There was an early burst release for both the acetate and PBS media within the first two hours following incubation, followed by a gradual release throughout the following 12 h (13 and 36% for pH 7.4 and pH 5.5, respectively) (Fig. 4). Then, within 20 days, a constant release was seen in both buffers ($20.2 \pm 0.5\%$ in pH 7.4 and $50.1 \pm 0.5\%$ in pH 5.5). The cumulative drug release phase for NPs over the course of 36 days reached $27.4 \pm 0.3\%$ and $60.1 \pm 0.6\%$ at pH 7.4 and pH 5.5, respectively. Overall, the cumulative Dox release was found to be about three times larger at pH 5.5 than at pH 7.4 (Fig. 4), which may have been caused by the protonated Dox's enhanced solubility at lower pH. In conclusion, the NPs seem to be suitable for reaching the lowest possible release in blood and the highest possible release in lysosome, which depends on the pH of media, drug diffusion, and matrix-erosion mechanisms.

By deacetylating chitin, chitosan is a naturally occurring, biocompatible, biodegradable, and low-toxic substance. Its lengthy chain, produced by the fusion of glycosidic linkages and 2-amino-2-deoxy-d-glucan, results in a positive charge that pulls the systems toward the negatively charged cell membrane [42]. As a result, IOMNPs coated with chitosan can exhibit mucoadhesive qualities and improve nanoparticle retention in the target regions [30]. Since chitosan coating does not alter the thermal or magnetic properties of IOMNPs, numerous systems were created

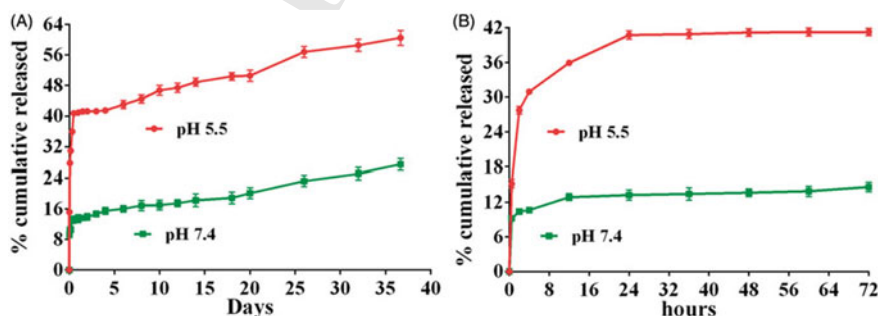


Fig. 4 In vitro release profile of Dox from NPs in pH 7.4 PBS and pH 5.5 acetate buffers. **a** For 36 days and **b** for 72 h (n = 3) [41]

242 [30]. Due to the electrostatic attraction between the positively charged nanoparticles,
243 chitosan with low molecular weight can prevent these nanoparticles from aggregating
244 [43]. Due to the partial protonation of its amino groups in water at physiological pH,
245 which decreases chitosan solubility, this polymer exhibits some limits as a coating
246 material. These issues can be resolved chemically, increasing the water solubility of
247 chitosan derivatives [44]. For the pH-responsive release of 5-FLU delivery in A549
248 and HeLa S3 cell cultures and the ensuing synergistic cytotoxicity of 5-FLU and
249 ROS, Ayyanaar et al. (2020) presented the synthesis, characterization, and evaluation
250 of chitosan-coated magnetic iron oxide nanoparticles ($\text{Fe}_3\text{O}_4@OA\text{-CS-5-FLU-NPs}$)
251 [45]. In this case, magnetic nanoparticles were created using the conventional co-
252 precipitation method. It was simple and quick to create $\text{Fe}_3\text{O}_4@OA\text{-CS-5-FLU-NPs}$
253 using in situ loading. The proposed $\text{Fe}_3\text{O}_4@OA\text{-CS-5-FLU-NPs}$ were successfully
254 synthesized and subjected to numerous spectroscopic and microscopic analyses to
255 characterize them. The $\text{Fe}_3\text{O}_4@OA\text{-CS-5-FLU-NPs}$ ' targeted drug release profile
256 was investigated in the presence of ROS, including H_2O_2 , and pH induction. The
257 magnetic characteristics of the $\text{Fe}_3\text{O}_4@OA\text{-CS-5-FLU-NPs}$ showed promise and
258 might be applied to magnetic targeted and pH-responsive drug delivery systems.
259 The ROS-responsive polymeric nanocarriers are released from $\text{Fe}_3\text{O}_4@OA\text{-CS-5-}$
260 FLU-NPs via pH-triggered drug release under slightly acidic conditions of pH 5.2
261 and pH 7.4. $\text{Fe}_3\text{O}_4@OA\text{-CS-5-FLU-NP}$, the product that was made available, revealed
262 satisfactory levels of cytotoxicity, morphological alterations, and inhibition of colony
263 formation for A549 and HeLa S3 cancer cells (Fig. 5). At 24 h, the IC_{50} values were
264 12.9 and 23 g/mL, respectively. The $\text{Fe}_3\text{O}_4@OA\text{-CS-5-FLU-NPs}$ were active and
265 safe for anticancer biomedical applications, according to the results of the MTT
266 assay, fluorescence staining, and colony formation assays. The system for enhanced
267 cancer theranostics presented by the current inquiry is strong and should be further
268 studied.

269 A modified magnetic nanoparticle was created and produced by Hami (2020) to be
270 employed as a targeted medicine delivery method [46]. On the surface of chitosan-
271 modified iron magnetic nanoparticles, nanocurcumin loading was done twice. In
272 the initial stage, chitosan was added as a coating polymer on the surface of iron
273 magnetic nanoparticles. The final reaction was carried out in the next step by coating
274 the surface of magnetic iron nanoparticles modified by chitosan with nanocurcumin.
275 The structure, shape, physicochemical properties, and presence of nanocurcumin
276 layers on chitosan in nanoparticles with a diameter of 20 nm were clearly seen in
277 the results of transmission and scanning electron microscopy. The iron–oxygen peak
278 in Fourier transform infrared spectroscopy showed magnetic iron nanoparticles, and
279 the oxygen–hydrogen peak revealed nanocurcumin layers on chitosan. Addition-
280 ally, X-ray spectroscopy demonstrated iron, carbon, oxygen, and nitrogen peaks,
281 which supported the presence of these components in the final composition. By
282 using a magnetic instrument called a vibration sample meter to analyze the magnetic
283 properties of iron-chitosan magnetic nanoparticles loaded with nanocurcumin, it
284 was discovered that the magnetic saturation of iron oxide nanoparticles at room
285 temperature was 63.1 emu g^{-1} as determined by the magnetic curve. Due to the

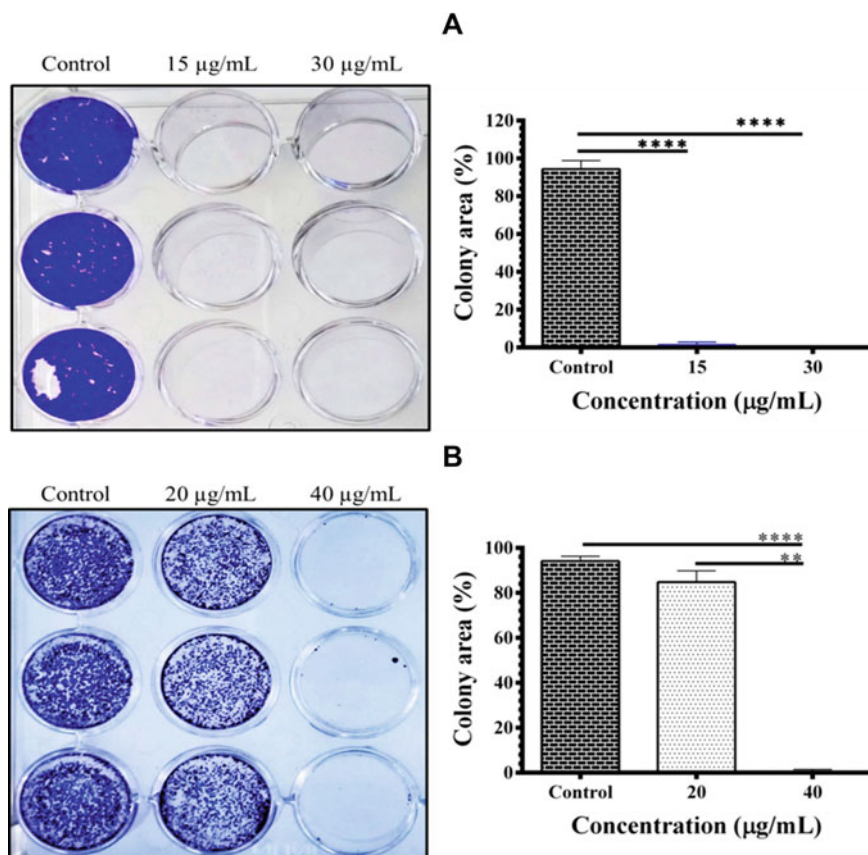


Fig. 5 Colony formation in the presence of $\text{Fe}_3\text{O}_4@OA\text{-CS-5-FLU-NP}$ examined against A549 and HeLa S3 cells: **a** A549 and **b** HeLa S3 cells. The colony area was calculated by ImageJ software using colony area (mean \pm SD). Significance of **** $p < 0.0001$ and ** $p < 0.01$ as compared with the untreated control group [45]

286 stabilization of polymeric groups on the nanoparticle surface, the magnetic saturation
 287 (35.3 emu g^{-1}) of iron-chitosan magnetic nanoparticles loaded with nanocurcumin clearly decreased. This proof showed that iron magnetic nanoparticles modified with chitosan successfully loaded with nanocurcumin. The results of this study
 288 showed that nanocurcumin was successfully loaded on the surface of magnetic iron
 289 nanoparticles modified by chitosan.
 290

291
 292 Inorganic substances, bioactive chemicals, and organic surfactants can also be
 293 applied on the surface of IOMNPs. Specifically, when IOMNPs are generated in
 294 organic solutions, organic surfactants are used to functionalize them. Dimercapto-
 295 succinic acid has the ability to produce nanoparticles with an anionic surface that
 296 resists opsonization and clearance by the reticuloendothelial system, hence lowering
 297 the toxicity to cells [30]. Additionally, oleic acid and trisodium citrate are able to

298 stabilize nanoparticles by generating repulsive forces (mostly steric repulsion) to
299 counterbalance the magnetic and van der Waals attractive forces [47]. In order to
300 achieve more effective coatings of the IOMNPs, with improved dispersion capacity in
301 solutions and lower nanoparticle clustering, surfactants showing lower values of critical
302 micelle concentrations were used [29]. This is because long hydrocarbon chains
303 in surfactants can produce hydrophobic nanoparticles. When utilized in IOMNP
304 systems, inorganic substances such as carbon, metals, silica, oxides (both metal and
305 non-metal), and sulfides showed the benefit of enhancing the antioxidant properties
306 of these nanoparticles [49]. SiO₂ can improve the dispersion of IOMNP in solutions,
307 increasing their stability and protecting them in acidic media [29].

308 A barrier against IOMNP oxidation, carbon-based coatings exhibit chemical and
309 thermal stability, high electrical conductivity, and solubility [30]. Additionally, the
310 creation of a positively charged silver coating by the electron transfer between silver
311 and IOMNPs in a nanosystem enables the conjugation of several antibiotics to the
312 silver-decorated nanoparticles [29]. IOMNPs frequently contain oxides and sulfides
313 to stabilize the nanosystem and promote desirable magnetic characteristics [30]. The
314 application of metal coatings with modifications including substances like thiol can
315 enable their coupling with a variety of biomolecules. ZnO was chosen as the best
316 compound for an anticancer nanosystem due to both its intrinsic anticancer properties
317 and biocompatibility because the choice of a coating for the IOMNPs must take into
318 account both their intrinsic properties and the purpose of the system [29]. Examples
319 of bioactive compounds that can be employed in IOMNP-based systems include
320 peptides, lipids, and proteins. They ought to be able to preserve both the magnetic
321 characteristics and the stability of the nanoparticles [30].

322 Concluding Remarks

323 This review summarizes the roles of untreated and modified iron oxide nanoparti-
324 cles for drug delivery. The superior size, surface, mechanical, optical, and magnetic
325 properties of iron oxide NPs make them ideal for drug delivery. The nanoparti-
326 cles' tiny size and wide surface area boost their solubility and bioavailability, which
327 in turn increases their capacity to penetrate the blood-brain barrier (BBB), enter
328 the respiratory system, and be absorbed through the skin's tight connections. The
329 majority of the studies under review functionalized the iron oxide NPs' surface with
330 targeting ligands, imaging, and therapeutic moieties. This allowed for the protection
331 of IOMNPs with biocompatible materials to stop biodegradation, changes, and aggre-
332 gation as well as the entrapment of the bioactive agent on the particle via adsorption
333 or covalent attachment. As a result of a stronger drug action at lower doses and a
334 consequent improvement in therapy, IOMNPs exhibit significant promise for use in
335 medication delivery. This will minimize toxicity and adverse effects. These results
336 show that IOMNPs may be used in drug delivery applications, and that achieving
337 adequate drug release control on the target tissues is beneficial in a variety of clinical

338 conditions, including infections, inflammations, and malignancies. However, given
339 their therapeutic uses, further toxicological clinical studies on IOMNPs are required.

340 References

- 341 1. Omorogbe SO, Aigbodion AI, Ifijen HI, Ogbeide-Ihama N, Simo A, Ikhuoria EU (2020) Low
342 temperature synthesis of super paramagnetic Fe₃O₄ morphologies tuned using oleic acid as
343 crystal growth modifier. In: Book TMS, 149th annual meeting & exhibition supplemental
344 proceedings, pp 619–631
- 345 2. Ifijen IH, Ikhuoria EU, Maliki M, Otabor GO, Aigbodion AI (2022) Nanostructured materials:
346 a review on its application in water treatment. In: The Minerals, Metals & Materials Society
347 (eds) TMS 2022 151st annual meeting & exhibition supplemental proceedings. The Minerals,
348 Metals & Material Series. Springer, Cham, pp 1172–1180
- 349 3. Ifijen IH, Aghedo ON, Odiachi IJ, Omorogbe SO, Olu EL, Onuguh IC (2022) Nanostructured
350 graphene thin films: a brief review of their fabrication techniques and corrosion protective
351 performance. In: The Minerals, Metals & Materials Society (eds) TMS 2022 151st annual
352 meeting & exhibition supplemental proceedings. The Minerals, Metals & Material Series.
353 Springer, Cham, pp 366–377
- 354 4. Ifijen IH, Maliki M, Omorogbe SO, Ibrahim SD (2022) Incorporation of metallic nanoparticles
355 into alkyd resin: a review of their coating performance. In: The Minerals, Metals & Materials
356 Society (eds) TMS 2022 151st annual meeting & exhibition supplemental proceedings. The
357 Minerals, Metals & Material Series. Springer, Cham, pp 338–349
- 358 5. Omorogbe SO, Ikhuoria SO, Ifijen IH, Simo A, Aigbodion AI, Maaza M (2019) Fabrication of
359 monodispersed needle-sized hollow core polystyrene microspheres. In: The Minerals, Metals &
360 Minerals Society (ed) TMS 2019 148th annual meeting & exhibition supplement proceeding,
361 pp 155–164
- 362 6. Ifijen IH, Ikhuoria EU (2019) Generation of highly ordered 3d vivid monochromatic coloured
363 photonic crystal films using evaporative induced technique. Tanzania J Sci 45(3):439449
- 364 7. Ifijen IH, Ikhuoria EU (2020) Monodisperse polystyrene microspheres: studies on the effects
365 of reaction parameters on particle diameter. Tanzania J Sci 46(1):19–30
- 366 8. Ifijen IH, Ikhuoria EU, Omorogbe SO (2018) Correlative studies on the fabrication of
367 poly (styrene-methyl-methacrylate-acrylic acid) colloidal crystal films. J Dispersion Sci Tech
368 40(7):1–8
- 369 9. Ifijen IH, Ikhuoria EU, Omorogbe SO, Aigbodion AI (2019a) Ordered colloidal crystals fabri-
370 cation and studies on the properties of poly (styrene-butyl acrylate-acrylic acid) and polystyrene
371 latexes. In: Srivatsan T, Gupta M (eds) Nanocomposites VI: nanoscience and nanotechnology
372 in advanced composites. The Minerals, Metals & Minerals Series. Springer, Cham, pp 155–164
- 373 10. Ifijen IH, Maliki M, Ovonramwen OB, Aigbodion AI, Ikhuoria EU (2019) Brilliant coloured
374 monochromatic photonic crystals films generation from poly (styrene-butyl acrylate-acrylic
375 acid) latex. J Appl Sci Environ Mgt 23(9):1661–1664
- 376 11. Ifijen IH, Omorogbe SO, Maliki M, Odiachi IJ, Aigbodion AI, Ikhuoria EU (2020) Stabilizing
377 capability of gum arabic on the synthesis of poly (styrene-methylmethacrylate-acrylic acid)
378 latex for the generation of colloidal crystal films. Tanzania J Sci 46(2):345–435
- 379 12. Ifijen IH, Maliki M, Odiachi IJ, Aghedo ON, Ohiocheoya EB (2022) Review on solvents-based
380 alkyd resins and water borne alkyd resins: impacts of modification on their coating properties.
381 Chem Afri 5:211–225
- 382 13. Ifijen IH, Ikhuoria EU, Aigbodion AI, Omorogbe SO (2018) Impact of varying the concentration
383 of tetraethyl-orthosilicate on the average particle diameter of monodisperse colloidal silica
384 spheres. Chem Sci J 9(1):183–185

- 385 14. Ikuhuria EU, Ifijen IH, Georgina OP, Ehigie AC, Omorogbe SO, Aigbodion AI (2020) The
386 adsorption of heavy metals from aqueous solutions using silica microparticles synthesized from
387 sodium silicate. In: Ni–Co 2021: the 5th international symposium on Ni and Co, pp 195–205
- 388 15. Ifijen IH, Itua AB, Maliki M, Ize-Iyamu CO, Omorogbe SO, Aigbodion AI, Ikuhuria EU
389 (2020) The removal of nickel and lead ions from aqueous solutions using green synthesized
390 silica microparticles. *Heliyon* 6(9):e04907
- 391 16. Ifijen IH, Maliki M (2022) A comprehensive review on the synthesis and photothermal cancer
392 therapy of titanium nitride nanostructures. *Inorg Nano-Metal Chem.* [https://doi.org/10.1080/
393 24701556.2022.2068596](https://doi.org/10.1080/24701556.2022.2068596)
- 394 17. Jonathan EM, Ifijen IH, Mokobia KE, Okeke EI, Omoruyi CI, Anegebe B (2022) A review on
395 the heightened mechanical features of nanosilica-based concrete and the response of human
396 fibroblasts to nanosilica. *Biomed Mater Dev* <https://doi.org/10.1007/s44174-022-00013-4>
- 397 18. Characterization of sulphated cellulose nanocrystals as stabilizer for magnetite nanoparticles
398 synthesis with improved magnetic properties. *Nig J Mater Sci Eng* 7(2):23–31
- 399 19. Omorogbe SO, Ikuhuria EU, Igiehon LI, Agbonlahor GO, Ifijen IH, Aigbodion AI (2017)
400 Characterization of sulphated cellulose nanocrystals as stabilizer for magnetite nanoparticles
401 synthesis with improved magnetic properties. *Nigerian J Mater Sci Technol* 7:21–30
- 402 20. Wu W, Wu W, Yu T, Jiang C, Kim W-K (2015) Recent progress on magnetic iron oxide
403 nanoparticles: synthesis, surface functional strategies and biomedical applications. *Sci Technol*
404 *Adv Mater* 16:2
- 405 21. Ali A, Hira Zafar MZ, UI I, Phull AR, Ali JS, Hussain A (2016) Synthesis, characterization,
406 applications, and challenges of iron oxide nanoparticles. *Nanotechnol Sci Appl* 9:49
- 407 22. Raza A, Hayat U, Rasheed T, Bilal M, Iqbal HM (2019) Smart materials-based near-infrared
408 light-responsive drug delivery systems for cancer treatment: a review. *J Mater Res Technol*
409 8:1497–1509
- 410 23. Pinelli F, Perale G, Rossi F (2020) Coating and functionalization strategies for nanogels and
411 nanoparticles for selective drug delivery. *Gels* 6:6
- 412 24. Vega-Vasquez P, Mosier NS, Irudayaraj J (2020) Nanoscale drug delivery systems: from
413 medicine to agriculture. *Front Bioeng Biotechnol* 8:79
- 414 25. Ebadi M, Buskaran K, Bullo S, Hussein MZ, Fakurazi S, Pastorin G (2020) Drug
415 delivery system based on magnetic iron oxide nanoparticles coated with (polyvinyl alcohol-
416 zinc/aluminium-layered double hydroxide-sorafenib). *Alexandria Engin J* 60:733–747
- 417 26. Laurent S, Bridot J-L, Elst LV, Muller RN (2010) Magnetic iron oxide nanoparticles for
418 biomedical applications. *Future Med Chem* 2:427–449
- 419 27. Pham X-H, Hahn E, Kim H-M, Son BS, Jo A, An J, Thi T, An T, Nguyen DQ, Jun B-H
420 (2010) Silica-coated magnetic iron oxide nanoparticles grafted onto graphene oxide for protein
421 isolation. *Nanomater* 10:117
- 422 28. Dulinska-Litewka J, Łazarczyk A, Hałubiec P, Szafranski O, Karnas K, Karewicz A (2019)
423 Superparamagnetic iron oxide nanoparticles—current and prospective medical applications.
424 *Mater* 12:617
- 425 29. Bruschi ML, de Toledo LAS (2019) Pharmaceutical applications of iron-oxide magnetic
426 nanoparticles. *Magnetochem* 5:50
- 427 30. Arias LS, Pessan JP, Vieira APM, De Lima TMT, Delbem ACB, Monteiro DR (2018) Iron oxide
428 nanoparticles for biomedical applications: a perspective on synthesis, drugs, antimicrobial
429 activity, and toxicity. *Antibiotics* 7:46
- 430 31. Hao X, Xu B, Chen H, Wang X, Zhang J, Guo R, Shi X, Cao X (2019) stem cell-mediated
431 delivery of nanogels loaded with ultrasmall iron oxide nanoparticles for enhanced tumor MR
432 imaging. *Nanoscale* 11:4904–4910
- 433 32. Turrina C, Berensmeier S, Schwaminger SP (2021) Bare iron oxide nanoparticles as drug
434 delivery carrier for the short cationic peptide lasioglossin. *Pharmaceuticals* 14:405
- 435 33. Kansara K, Patel P, Shukla RK, Pandya A, Shanker R, Kumar A, Dhawan A (2018) Synthesis
436 of biocompatible iron oxide nanoparticles as a drug delivery vehicle. *Int J Nanomed* 13:79–82
- 437 34. Laurent S, Forge D, Port M, Roch A, Robic C, Elst LV, Muller RN (2008) Magnetic iron oxide
438 nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and
439 biological applications. *Chem Rev* 108:2064–2110

- 440 35. Li F, Liang Z, Liu J, Sun J, Hu X, Zhao M, Liu J, Bai R, Kim D, Sun X et al (2019) Dynamically reversible iron oxide nanoparticle assemblies for targeted amplification of T1-weighted
441 magnetic resonance imaging of tumors. *Nano Lett* 19:4213–4220
- 442 36. Yallapu MM, Foy SP, Jain TK, Labhasetwar V (2010) PEG-functionalized magnetic nanoparticles
443 for drug delivery and magnetic resonance imaging applications. *Pharm Res* 27(11):2283–
444 2295
- 445 37. Mohanta SC, Saha A, Devi PS (2018) PEGylated iron oxide nanoparticles for pH responsive
446 drug delivery application. *Mater Today Proceed* 5(3):9715–9725
- 447 38. Bruschi ML (2015) Strategies to modify the drug release from pharmaceutical systems. Elsevier,
448 Amsterdam, The Netherlands
- 449 39. Butoescu N, Jordan O, Burdet P, Stadelmann P, Petri-Fink A, Hofmann H, Doelker E, Fink A
450 (2009) Dexamethasone-containing biodegradable superparamagnetic microparticles for intra-
451 articular administration: Physicochemical and magnetic properties, in vitro and in vivo drug
452 release. *Eur J Pharm Biopharm* 72:529–538
- 453 40. Hajikarimi Z, Khoei S, Khoee S, Mahdavi SR (2014) Evaluation of the cytotoxic effects of
454 PLGA coated iron oxide nanoparticles as a carrier of 5-fluorouracil and mega-voltage x-ray
455 radiation in DU145 prostate cancer cell line. *IEEE Trans Nanobiosci* 13(4):403–408
- 456 41. Mosafer J, Teymouri M (2018) Comparative study of superparamagnetic iron
457 oxide/doxorubicin co-loaded poly (lactic-co-glycolic acid) nanospheres prepared by different
458 emulsion solvent evaporation method. *Artif Cells Nanomed Biotechnol* 46(6):1146–1155
- 459 42. Banerjee T, Mitra S, Singh AK, Sharma RK, Maitra A (2002) Preparation, characterization
460 and biodistribution of ultrafine chitosan nanoparticles. *Int J Pharm* 243:93–105
- 461 43. Parsian M, Unsoy G, Mutlu P, Yalcin S, Tezcaner A, Gündüz U (2016) Loading of Gemcitabine
462 on chitosan magnetic nanoparticles increases the anti-cancer efficacy of the drug. *Eur J*
463 *Pharmacol* 784:121–128
- 464 44. Saikia C, Hussain A, Ramteke A, Sharma HK, Maji TK (2015) Carboxymethyl starch-chitosan-
465 coated
466 Iron Oxide Magnetic Nanoparticles for Controlled Delivery of Isoniazid. *J Microencapsul*
467 32:29–39
- 468 46. Ayyanaar S, Balachandran C, Bhaskar RC, Kesavan MP, Aoki S, Raja RP, Rajesh J, Webster
469 TJ, Rajagopal G (2020) ROS-responsive chitosan coated magnetic iron oxide nanoparticles as
470 potential vehicles for targeted drug delivery in cancer therapy. *Int J Nanomed* 15:3333–3346
- 471 47. Hami Z (2020) Coating iron oxide nanoparticles with chitosan for targeted delivery of
472 nanocurcumin. *Ann Mil Health Sci Res* 18(1):e103657
- 473 48. Soares P, Lochte F, Echeverria C, Pereira LC, Coutinho JT, Ferreira IM, Novo CM, Borges
474 JP (2015) Thermal and magnetic properties of iron oxide colloids: influence of surfactants.
475 *Nanotechnol* 26:425704
- 476 49. Luchini A, Heenan RK, Paduano L, Vitiello G (2016) Functionalized SPIONs: the surfactant
477 nature modulates the self-assembly and cluster formation. *Phys Chem Chem Phys* 18:18441–
478 18449
- 479 50. El-Boubbou K (2018) Magnetic iron oxide nanoparticles as drug carriers: clinical relevance.
480 *Nanomed* 13:953–971
481

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Chapter 22

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